



Review

Nitric Oxide and Biological Mediators in Pediatric Chronic Rhinosinusitis and Asthma

Valentina Agnese Ferraro *, Stefania Zanconato, Eugenio Baraldi and Silvia Carraro

Women's and Children's Health Department, University of Padova, via Giustiniani 2, 35128 Padova, Italy; stefania.zanconato@aopd.veneto.it (S.Z.); eugenio.baraldi@unipd.it (E.B.); silvia.carraro@unipd.it (S.C.)

* Correspondence: ferrarovalentina@hotmail.com; Tel.: +39-0498213505

Received: 7 September 2019; Accepted: 23 October 2019; Published: 25 October 2019



Abstract: Background: In the context of the so-called unified airway theory, chronic rhinosinusitis (CRS) and asthma may coexist. The inflammation underlying these conditions can be studied through the aid of biomarkers. Main body: We described the main biological mediators that have been studied in pediatric CRS and asthma, and, according to the available literature, we reported their potential role in the diagnosis and management of these conditions. As for CRS, we discussed the studies that investigated nasal nitric oxide (nNO), pendrin, and periostin. As for asthma, we discussed the role of fractional exhaled nitric oxide (feNO), the role of periostin, and that of biological mediators measured in exhaled breath condensate (EBC) and exhaled air (volatile organic compounds, VOCs). Conclusion: Among non-invasive biomarkers, nNO seems the most informative in CRS and feNO in asthma. Other biological mediators seem promising, but further studies are needed before they can be applied in clinical practice.

Keywords: nitric oxide; biological mediators; pediatric rhinosinusitis; pediatric asthma

1. Introduction

The possible coexistence of rhinosinusitis and asthma is well known [1,2], in the context of the so-called unified airway theory, which describes the upper and lower airways as a single functional unit [3–5]. Moreover, the upper and lower respiratory tracts have common histological structures, including the basement membrane, lamina propria, ciliary epithelium, glands, and goblet cells [6].

Nowadays, more and more interest is directed to the study of noninvasive tests, which could help assess the presence and nature of airway inflammation in childhood chronic rhinosinusitis (CRS) and asthma, in order to learn more about the underlying pathological pathways of these complex diseases, and potentially guiding the development of a personalized medicine.

The better studied noninvasive marker of airway inflammation is nitric oxide (NO). NO is a free radical gas produced from L-arginine mainly by two enzymes: constitutive nitric oxide synthase (NOS), which constantly generates low concentrations of NO, and inducible NOS (iNOS), also called type 2 NOS, which is present in airway epithelial cells, where it is upregulated by proinflammatory cytokines, such as tumor necrosis factor and interleukin-1 β , and by lipopolysaccharides of Gram-negative bacteria [7–9].

NO is released throughout the airways, and both the NO released from the upper respiratory tract (nasal NO, nNO) and the lower respiratory tract (fractional exhaled nitric oxide, feNO) can be measured.

Nasal NO can be measured with a non-invasive method based on the nasal aspiration, at a constant flow rate, from one naris with gas entrained via the other naris (transnasal flow in series) during velum closure, in order to prevent leak of nasal NO via the posterior velopharyngeal aperture and to reduce contamination of nasal gas with lower airway air [10,11]. Till now, no other recommended methods

have been described, even if this method requires patient collaboration, and alternatives have been studied for children who cannot manage velum closure [12–15].

The standardized method to measure feNO is the single breath on-line (SBOL) method, which is non-invasive, rapid, repeatable, and reproducible. The subject has to inhale through the mouth to total lung capacity (TLC), then exhaling during velum closure (against a positive pressure of 5–20 cmH₂O) [11,16]. This technique is well standardized in children who are able to cooperate, while until now, no standardized methods have been recommended in young or uncooperative children [17–19]. In these children, the most appropriate method is the tidal breathing offline method, which is carried out collecting exhaled air in an appropriate reservoir for later analysis [20–22]. One limit of this method is the lack of control in the expiratory flow, being feNO values highly flow-dependent. To overcome this problem, fast-response chemiluminescence analyzers and flow control devices have been developed [23], as well as mathematical algorithms, which try to obtain, from tidal breathing feNO values, the corresponding single breath flow values [24]. The second limit of the tidal breathing offline method is the contamination from nose-derived air, which contains higher NO levels that may affect the measurements of feNO; for this reason, the use of facemasks with a septum that separates the air from upper and lower airways has been suggested [22].

Here, we discussed the possible role of NO as a biomarker in chronic rhinosinusitis (CRS) and asthma in children. Moreover, we described the main other biological mediators that have been evaluated in these conditions. In particular, the possible role of periostin and pendrin was reported, and the potential of molecules measured in exhaled air (volatile organic compounds, VOCs) and exhaled breath condensate was discussed. Figure 1 summarizes these biomarkers, highlighting the current applicability in clinical practice.

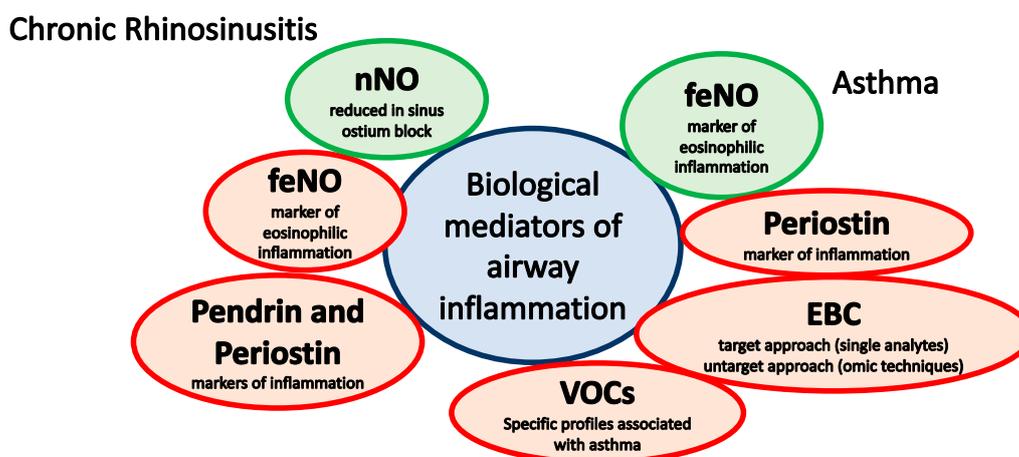


Figure 1. Biological mediators in pediatric rhinosinusitis and asthma. Green: applicable to clinical practice; Red: not applicable to clinical practice yet. nNo: nasal nitric oxide; feNO: fractional exhaled nitric oxide; EBC: exhaled breath condensate; VOCs: volatile organic compounds.

2. Chronic Rhinosinusitis

Even if the diagnosis of CRS in children is based on symptoms (nasal obstruction, nasal discharge, facial pain/pressure, and reduced/lost smell) persisting for more than 12 weeks with associated endoscopic or radiographic findings [25], a number of biomarkers has been studied for their possible role in diagnosis, work-up, and management of this condition [2].

2.1. Nitric Oxide

NO has been extensively studied in CRS, mainly because upper airways represent the main source of respiratory NO [26,27], and, additionally, paranasal sinuses are the main production site [28]. In the upper airways, NO has several functions: a specific host defense against infective agents (bacteria,

viruses, and fungi) [29], modulator of cilia motility [30], a regulator of nasal airflow humidification and warming [31], airborne messenger between higher and lower respiratory tracts [32,33].

The role of nasal NO in sinusitis has been evaluated in some studies. The first study describing variations of nasal NO levels during acute sinusitis was published in 1997 by Baraldi et al. [34]. The authors showed that children with acute maxillary sinusitis had a marked reduction in nasal NO concentrations, and nNO returned to normal levels after antibiotic therapy. This finding could be caused by mechanical obstruction of the draining ostia and by negative pressure within the sinuses, resulting in a decreased passage of NO from sinuses to the nasal lumen [34]. In keeping with this, in adult patients, it was demonstrated that despite patients with CRS with nasal polyps had high level of NOS2 in nasal epithelium, because of the inflammation of nasal and paranasal cavities, they exhibited lower nNO compared to patients with uncomplicated allergic rhinitis; moreover, the higher was the extent of polyposis and the lower were the levels of nNO [35]. This was probably due to the osteomeatal complex obstruction, which was associated with the inability of NO produced in the sinuses to reach the nasal cavity [35]. Likewise, another study confirmed, in patients with polyposis (which lead to blockade of the osteomeatal complex), a negative correlation between nNO and the degree of sinus disease [36]. In keeping with this, in a patient with CRS and polyposis, nNO diminished with the increase of sinus opacification observed with CT scans [37]. Even if these studies were mostly performed in adults (Table 1 summarizes these studies), they pointed out that inflammation of sino-nasal mucosa, especially if associated with polyps, prevents NO to flow from sinuses to the nasal lumen, so that reduced nNO levels are measured in these conditions.

Noteworthy, the reduction of nasal NO may lead to a vicious circle, increasing the risk of recurrent infections in these subjects, since NO plays a role in host airway defenses against exogenous agents [38].

In conclusion, although the clinical relevance of finding reduced nNO levels in CRS is limited [8,39], this biomarker can be useful to monitor sinus ostium block during both post-medical and postsurgery follow-up in subjects affected by bilateral nasal polyposis [8].

Table 1. List of studies that evaluated nasal nitric oxide (nNO) in rhinosinusitis.

Study	Aims	Population	Results (mean ± SD)	Conclusions
[34]	To evaluate nNO in children with acute maxillary sinusitis before and after treatment with antibiotic therapy	16 children (4–13 years) with acute maxillary sinusitis; 16 age- and sex-matched healthy control subjects	(1) nNO = 70 ± 8.7 ppb sinusitis before antibiotic therapy; (2) nNO = 220 ± 15 ppb sinusitis after antibiotic therapy (amoxicillin/clavulanate); (3) nNO = 245 ± 15 ppb healthy control subjects	During acute maxillary sinusitis, nNO is decreased; nNO returns to normal after antibiotic therapy
[37]	To examine if nNO is affected by paranasal sinus inflammatory diseases	20 patients with nonallergic nasal polyposis (age 48 ± 3 years); 42 control subjects (age 42 ± 3 years)	(1) nNO = 150 ± 20 ppb in patients with nasal nonallergic polyposis; (2) nNO = 223 ± 6 ppb in controls	nNO in patients with nasal polyposis is decreased compared to controls, and it depends on the degree of obstruction of the paranasal sinuses
[35]	To evaluate nNO in patients with nasal polyposis compared with allergic rhinitis and to analyze the effect of polyp treatment on nNO	44 patients with rhinitis without polyps (age = 39 ± 13.6 years) and 38 with polyps (age = 45.6 ± 4.5 years); 20 normal controls (age = 36.9 ± 11.6 years); 23 patients with polyposis pre- and post-treatment (age = 48.8 ± 4.2 years)	(1) nNO = 740.9 ± 148.1 ppb in normal controls (2) nNO = 659.8 ± 304.8 ppb in allergic rhinitis (3) nNO is significantly lower in patients with polyps than allergic rhinitis without polyps (Kruskal–Wallis, $p = 0.0001$, $\chi^2 = 37.6$, d.f. = 4) (4) Successful treatment, with reduction in polyp volume, associated with a rise in NO levels ($p = 0.042$)	nNO levels are low in nasal polyps. A rise in nNO is seen with successful polyp treatment

Table 1. Cont.

Study	Aims	Population	Results (mean ± SD)	Conclusions
[36]	To study the effect of CRS therapy on nNO and to see whether nNO changes correlate with other assessments.	90 patients (mean age 43 ± 13 years) with CRS who still had troublesome symptoms after initial therapy with dexarhinaspary and nasal douching	(1) Baseline nNO correlate with CT scores ($p < 0.001$) (2) The mean nNO levels for the three grades of severity at CT scan are grade1 (less severe) 537 ± 202 , grade 2 362 ± 188 , and grade 3 165 ± 151 ppb. (3) The percentage rise in nNO correlates with changes in symptom scores ($p < 0.001$), saccharin clearance time ($p < 0.001$), endoscopic changes ($p < 0.001$), polyp grades ($p < 0.05$ at 6 months, $p < 0.01$ at 12 months), and surgical scores ($p < 0.01$).	nNO provides a valuable non-invasive objective measure of the response of CRS to therapy

ppb: parts per billion; CRS: chronic rhinosinusitis; nNO: nasal nitric oxide.

Even if feNO is considered a marker of lower airway eosinophilic inflammation, in the context of the previously described unified airway theory, recent studies have analyzed this marker in CRS, with variable results. Kobayashi et al. showed that patients with eosinophilic CRS without asthma did not have high feNO levels, while feNO levels were elevated in well-controlled asthmatic patients with eosinophilic CRS [40]. On the other hand, Zhang et al. showed that 29% of the patients with CRS with nasal polyps, without pulmonary disease, had increased feNO [41], and Jeong et al. showed that 30 non-asthmatic, non-atopic patients with CRS with nasal polyps had a significantly higher feNO than healthy controls [42]. Similarly, Takeno et al. analyzed 33 patients with CRS with nasal polyps and found high feNO (defined as feNO >25 ppb) in 22 (66%), 8 (36%) of which with no history of asthma [43]. Furthermore, recent studies demonstrated that in patients with eosinophilic CRS, feNO levels correlated with the severity of the CT findings [40,44], and a reduction of this biomarker had been described after functional endoscopic sinus surgery [44].

In conclusion, the available data on feNO levels in CRS is not unanimous, and for the time being, there are no clear recommendations for its clinical use.

2.2. Pendrin and Periostin

Pendrin is an ion exchanger involved in inflammation and mucus production in patients with CRS, as well as in asthmatic patients [45,46]. Its role in mucus production is not only due to a direct effect but also mediated by the recruitment of inflammatory cells [46]. Pendrin has also been shown to regulate epithelial air-surface liquid levels and composition [45,47].

It has been demonstrated that pendrin is overexpressed in the sinonasal tissue, including epithelial cells and submucosal gland cells, in patients with CRS and nasal polyps, suggesting a pathogenetic role for this molecule [48]. The increased levels of pendrin might contribute to chronic inflammatory response, mucus production, and decreased mucociliary clearance [49].

Periostin is an extracellular matrix protein, a highly inducible product of IL-4 or IL-13, which are signature cytokines of the Th2-type immune response [50,51]. Moreover, periostin plays an important role as a regulator of fibrosis and collagen deposition [52]. Recently, the analysis of sinonasal mucosal biopsies, obtained from CRS patients, showed that periostin was associated with the presence of basement membrane thickening, fibrosis, and tissue eosinophilia and might identify patients undergoing remodeling changes [53]. In addition, its overproduction in the nasal mucosa of patients with CRS has been suggested to contribute to polyp formation [48,54,55]. Xu et al. demonstrated that periostin and VEGF (vascular endothelial growth factor) were higher in eosinophilic nasal polyps than in non-eosinophilic nasal polyps and control tissue, and in vitro VEGF was upregulated by periostin, suggesting that periostin might play an important role in the development of eosinophilic nasal polyps [56]. It has been shown that serum periostin in combination with blood eosinophils and basophils count has the potential to discriminate eosinophilic nasal polyps and non-eosinophilic nasal

polyps [57] and, in combination with IgE and *Staphylococcal enterotoxin* (SE)-IgE, may be useful to identify nasal polyps with moderate and severe type 2 inflammation [58].

In conclusion, the analysis of pendrin and periostin can provide some insights into the pathogenetic mechanisms involved in CRS. Nonetheless, such measurements are still limited to research, and they do not have a role in clinical practice yet.

3. Asthma

In the past two decades, many studies in the field of asthma focused on the investigation of biomarkers relevant for the diagnosis, phenotyping, or treatment of the disease [59–61]. Here, we discussed the most widely studied biomarkers, focusing on those measured non-invasively, since this aspect is particularly important when dealing with children

3.1. Fractional Concentration of Exhaled Nitric Oxide

Four years after the first report on the presence of nitric oxide in exhaled human breath [62], an increase in its levels has been reported in children with asthma [63], in particular during asthma exacerbation with a rapid decline after oral steroid therapy [64]. Subsequently, several studies analyzed feNO in pediatric asthma, demonstrating its role as a marker of eosinophilic airway inflammation, since it is correlated with eosinophil counts in blood and induced sputum or bronchoalveolar lavage fluid, with serum eosinophil cationic protein and with IgE levels [9,65–67]. Recently, it has been described a fair diagnostic accuracy of feNO for identifying asthmatic patients [68] and, as recently suggested by Pavord et al., for identifying the treatable trait of eosinophilic asthma (e.g., to identify patients who are likely to benefit from inhaled corticosteroids) [61]. On the other hand, being feNO increased also in other atopic conditions, other authors suggested that low feNO levels predict a non-eosinophilic asthma phenotype better than high levels can predict an eosinophilic one [69,70].

As far as it concerns young wheezing children, several studies demonstrated that feNO levels were higher in those with recurrent wheezing compared to healthy controls [71–73], in those with frequent wheezing with high asthma predictive index (API) compared to low API [74,75], and in those with persistent wheezing compared to transient wheezing [76]. Therefore, it has been suggested that feNO could help phenotype preschool-age children with recurrent wheezing, contributing to the identification of those with early-onset asthma. In detail, it has been demonstrated that in high-risk preschool children (at a mean age of 22 months), high feNO levels were associated with increased risk for school-age asthma [77], and in preschool children with symptoms suggestive of asthma, both feNO and specific IgE to inhalant allergens were associated with asthma at 8 years [78]. In addition, in longitudinal cohort studies in infants and toddlers (<2 years) with recurrent wheezing, feNO values higher than or equal to 30 ppb had a high predictive value for persistent wheezing at 3 years of age [79], and an increase in feNO was associated with a decrease in lung function 6 months later [80].

The predictive value of feNO for the development of asthma was analyzed even in healthy school children, showing that children in the highest feNO quartile had an increased risk of developing asthma compared to those with the lowest quartile [81]. Also, a study carried out in children (mean age 8.4 years, follow-up 5 years), with allergic rhinitis (without asthma) and feNO > 35 ppb at baseline, demonstrated a higher risk of new-onset asthma and a higher decrease in lung function, suggesting less lung growth in children with high feNO values [82].

feNO is also a marker of inhaled corticosteroids responsiveness, as well as a possible marker of treatment compliance since several studies demonstrated a drop in its levels in response to steroid therapy [71,83,84]. A recent systematic review and meta-analysis showed that adjusting treatment, according to feNO levels, reduced the likelihood of asthma exacerbations at the expense of increased inhaled corticosteroids doses [85].

In conclusion, feNO has been studied in particular for its potential role in eosinophilic asthma detection, early asthma identification, and corticosteroid responsiveness prediction. These possible clinical applications are well summarized in the available international guidelines (Table 2):

- The National Institute for Health and Care Excellence (NICE, 2017) recommends measuring feNO (described as positive test when more than or equal to 35 ppb) in children (aged 5 to 16 years) with symptoms suggestive of asthma, if there is diagnostic uncertainty after initial assessment (e.g., normal spirometry or obstructive spirometry with a negative bronchodilator reversibility test) [86]. Furthermore, using feNO to monitor asthma control is not routinely recommended [86].
- The 2019 British Thoracic Society guidelines recommend to use feNO (if available) to find evidence of eosinophilic inflammation (regard a feNO level of 35 ppb or more as a positive test), keeping in mind that a positive test increases the probability of asthma, but a negative test does not exclude asthma [87]. Also, except in specialist asthma clinics, the routine use of feNO testing to monitor asthma in children is not recommended [87].
- The 2019 Global Initiative of Asthma (GINA) guidelines report that feNO is not useful for ruling in or ruling out a diagnosis of asthma nor for guiding asthma treatment in the general population. Among alternative strategies for adjusting asthma treatment in children, GINA guidelines report that feNO-guided treatment significantly reduces exacerbation rates compared with guidelines-based treatment (Evidence A). Furthermore, feNO seems to be a useful adjunct in diagnosing asthma in pre-school children with recurrent wheezing, in whom an elevated feNO (recorded 4 weeks from any URTI) predicts asthma at school age [88].

Table 2. How guidelines consider the use of fractional exhaled nitric oxide (feNO) as a biomarker of asthma ().

Guideline	Cut-off Value	How to Use feNO in Clinical Practice
[86]	feNO positive if more than or equal to 35 ppb	<ul style="list-style-type: none"> • in children (aged 5 to 16 years) with symptoms suggestive of asthma, if there is diagnostic uncertainty after the initial assessment • not routinely recommended to monitor asthma control
[87]	feNO positive if more than or equal to 35 ppb	<ul style="list-style-type: none"> • (if available) to find evidence of eosinophilic inflammation • a positive test increases the probability of asthma, but a negative test does not exclude asthma • the routine use of feNO testing to monitor asthma in children is not recommended, except in specialist asthma clinics
[88]	No clear cut-off value	<ul style="list-style-type: none"> • feNO is not useful for ruling in or ruling out a diagnosis of asthma • feNO is not useful for guiding asthma treatment in the general population, even if among alternative strategies for adjusting asthma treatment in children; feNO-guided treatment significantly reduces exacerbation rates compared with guidelines-based treatment (Evidence A)

feNO = fractional exhaled nitric oxide; ppb = parts per billion.

3.2. Periostin

As previously described, periostin is an extracellular matrix protein upregulated by classic type 2 cytokines IL-4 and IL-13 [50,51], which was described in several reports as a useful biomarker of T2-inflammation in adult asthmatic patients [89–91]. Moreover, in asthmatic patients, periostin plays an important role as a regulator of fibrosis, airway remodeling, collagen deposition, and mucus production from goblet cells [50,52,92,93].

Despite an increased periostin level described in children with asthma [94,95], periostin is unlikely to be a useful biomarker of type 2 inflammation in children, mainly because its levels increase due to bone growth and this may overlap with local production within the airways [93,96].

In conclusion, as in CRS, also in asthma, the study of periostin is currently limited to the field of research.

3.3. Exhaled Breath Condensate (EBC)

Among non-invasive methods for studying airway inflammation, exhaled breath condensate (EBC) is one of the most attractive. It is a biofluid collected during tidal breathing by cooling exhaled air by contact with a cold surface or condenser [83,97]. The condensate contains unstable volatile and semi- and non-volatile molecules, and its composition is thought to mirror that of the airway lining fluid; EBC is considered a promising biofluid, which allows the noninvasive study of pulmonary biochemical and inflammatory processes [83,98–100]. Many studies have investigated the possible role of EBC analysis in asthma, using both targeted (a measurement of single analytes) and untargeted (omic techniques) approaches.

Through a targeted approach, many single mediators related to inflammation and oxidative stress have been searched in EBC. Among them, the most relevant are:

- pH, which tended to be lower in children with severe or acute asthma but not in mild and stable disease [101–103];
- Leukotrienes (LT): LTB₄, a potent inflammatory mediator and a chemoattractant for neutrophils, was increased in the EBC of asthmatic children, being twice as high in steroid-naïve patients with asthma as in healthy subjects [104,105]; Cysteinyl leukotrienes (LTC₄, LTD₄, and LTE₄), powerful constrictors and proinflammatory mediators, were increased in particular in unstable or severe asthma [106–108];
- 8-isoprostane, hydrogen peroxide (H₂O₂), and other markers of oxidative stress, which were increased in asthma [107,109]; in particular, H₂O₂ correlated with disease severity, disease control, and response to steroid treatment [110,111];
- 3-nitrotyrosine (3-NT) and other nitric oxide metabolites that were more concentrated in the EBC of asthmatic children than in healthy controls [102,112,113].

Since no single biomarker can fully describe the pathogenic processes underlying complex chronic diseases, “-omic approaches” have been applied to study the overall biochemical-metabolic composition of exhaled breath condensate, with the potential for identifying analyte profiles characteristic of specific conditions [98,99,114,115]. Both proteomics and metabolomics have been applied to EBC in asthma research.

Proteomics is defined as the study of the complete assessment of proteins in a biological sample in order to identify potential biomarkers associated with a specific disease [114]; therefore, detecting distinct protein biomarkers in different pathologies may assist in disease diagnosis, monitoring, treatment, and prognosis [116]. The complexity of proteomics, due to alternative splicing, posttranscriptional, and translational modifications and the enormous dynamic range of protein concentrations in biological samples, makes this research field one of the most interesting in the last years, even if still an object of speculation [117]. As far as it concerns healthy subjects, several studies explored EBC in order to characterize their protein composition (proteome maps), which could be useful for future clinical studies dedicated to the discovery of novel protein biomarkers for pulmonary diseases [118,119]. In keeping with this, Bloemen et al. found a specific pattern of expressed peptides in asthmatic children [120].

Metabolomics, without any priori hypothesis, studies the metabolite composition (or metabolome) of a biological sample, using a spectroscopic technique (usually NMR spectroscopy and mass spectrometry). It is nowadays considered the “-omic” science that comes closer to phenotype expression because the metabolome is the result of both genetic influences and environmental stimuli [121,122]. Therefore, metabolomics provides a snapshot of the overall physiology of the host and its response to the environment [121]. In the EBC of asthmatic children, the metabolomic analysis was applied to characterize the airway biochemical fingerprints, enabling the discrimination of children with and without asthma [123]. Furthermore, in children with asthma, EBC metabolomic analysis distinguished different asthma phenotypes and enabled the identification of a specific profile associated with severe asthma [124].

In conclusion, the analysis of EBC, both using targeted and untargeted approaches, seems promising for the study of physio-pathological mechanisms underlying asthma. Nonetheless, despite two comprehensive Task Force reports of the European Respiratory Society (ERS) and American Thoracic Society (ATS) published in 2005 [100] and in 2017 [83], EBC-analysis is not yet routinely applicable in clinical setting, mainly, because of the poor reproducibility of biomarkers and the absence of large surveys for determination of reference-normal values [125]. In keeping with this, a recent review published by Bannier et al. showed that studies on EBC research in pediatric asthma, performed between 2013 and 2018, are hardly comparable due to large heterogeneity in study populations, study methods, EBC collection methodologies, EBC biomarkers, analytical methods, and limits of detection [126].

3.4. Volatile Organic Compounds (VOCs)

In the last decades, exhaled breath has been studied using a metabolomic approach in the so-called “breathome”, that is the fingerprint of volatile organic compounds (VOCs) [127]. Airway VOCs are organic chemicals (e.g., a chemical compound that contains carbon) originated from the upper and lower airways and also from the capillary bed near the alveoli [128]. In order to collect VOCs from exhaled breath, different methodological approaches have been studied, taking care to exclude organic compounds from ambient air, to apply the correct breathing maneuvers, and to use the most suitable sampling materials [128].

Two different techniques have been used to study exhaled VOC profiles: (a) gas chromatography with mass spectrometry, a quantitative method that identifies individual components, and (b) the electronic Nose (e-Nose), a qualitative method that obtains probabilistic discrimination between biomarker profiles [129,130].

In pediatric asthma, different VOCs profiles have been described in children with and without asthma [131–135]. The analysis of exhaled VOCs may contribute to asthma diagnosis [136] and the discrimination of asthmatic children from those with transient wheezing symptoms [137]. Also, several studies demonstrated the potential role of VOCs analysis in the prediction of asthma exacerbation [138,139] and in the characterization of children with persistently controlled and uncontrolled asthma [140]. On the contrary, recently, Bannier et al. reported that Aeonose (an easy-to-use hand-held eNose) used in children ≥ 6 years had high feasibility ($>98\%$ successful measurements), but a modest diagnostic accuracy for the discrimination between asthma and healthy controls [141].

In conclusion, VOCs analysis is an attractive non-invasive method that could contribute to the identification of asthmatic subjects, even if larger studies are needed, in order to standardize the procedures and validate the technique of sampling and analysis.

4. Conclusions

Several non-invasive biomarkers have been investigated to study inflammation in CRS and asthma. As far as it concerns CRS, there is clear evidence that in subjects with bilateral nasal polyposis, nNO is reduced because of sinus ostium block; other biomarkers have been studied in this condition (in particular, pendrin and periostin), but, nowadays, they have no clear role in clinical practice.

In pediatric asthma, feNO levels may have a role in the characterization of Th2-mediated eosinophilic inflammation in the early identification of asthma in pre-school children with recurrent wheezing and the prediction of steroid responsiveness. Nonetheless, the use of feNO measurements in clinical practice is still limited, as specified by the current international guidelines.

Eventually, even if several studies investigated the possible role of EBC and VOCs analysis in pediatric asthma, they are not ready for clinical practice yet, and larger studies are needed to standardize the procedures of sampling and analysis.

Author Contributions: Conceptualization and methodology, V.A.F., S.C., S.Z., E.B.; writing — original draft preparation, V.A.F.; review and editing, supervision, S.C., S.Z., E.B.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Rosati, M.G.; Peters, A.T. Relationships among allergic rhinitis, asthma, and chronic rhinosinusitis. *Am. J. Rhinol. Allergy* **2016**, *30*, 44–47. [[CrossRef](#)] [[PubMed](#)]
2. Massoth, L.; Anderson, C.; McKinney, K.A. Asthma and Chronic Rhinosinusitis: Diagnosis and Medical Management. *Med. Sci.* **2019**, *7*, 53. [[CrossRef](#)] [[PubMed](#)]
3. Licari, A.; Castagnoli, R.; Denicolò, C.F.; Rossini, L.; Marseglia, A.; Marseglia, G.L. The Nose and the Lung: United Airway Disease? *Front. Pediatr.* **2017**, *5*, 44. [[CrossRef](#)] [[PubMed](#)]
4. Stachler, R.J. Comorbidities of asthma and the unified airway. *Int. Forum Allergy Rhinol.* **2015**, *5* (Suppl. 1), S17–S22. [[CrossRef](#)] [[PubMed](#)]
5. Grossman, J. One airway, one disease. *Chest* **1997**, *111* (Suppl. 2), 11S–16S. [[CrossRef](#)]
6. Licari, A.; Caimmi, S.; Bosa, L.; Marseglia, A.; Marseglia, G.L.; Caimmi, D. Rhinosinusitis and asthma: A very long engagement. *Int. J. Immunopathol. Pharmacol.* **2014**, *27*, 499–508. [[CrossRef](#)]
7. Maniscalco, M.; Sofia, M.; Pelaia, G. Nitric oxide in upper airways inflammatory diseases. *Inflamm. Res.* **2007**, *56*, 58–69. [[CrossRef](#)]
8. Maniscalco, M.; Bianco, A.; Mazzarella, G.; Motta, A. Recent Advances on Nitric Oxide in the Upper Airways. *Curr. Med. Chem.* **2016**, *23*, 2736–2745. [[CrossRef](#)]
9. Kim, H.B.; Eckel, S.P.; Kim, J.H.; Gilliland, F.D. Exhaled NO: Determinants and Clinical Application in Children with Allergic Airway Disease. *Allergy Asthma Immunol. Res.* **2016**, *8*, 12–21. [[CrossRef](#)]
10. Lucas, J.S.; Walker, W.T. NO way! Nasal nitric oxide measurement in infants. *Eur. Respir. J.* **2018**, *51*, 1800958. [[CrossRef](#)]
11. American Thoracic Society; European Respiratory Society. ATS/ERS recommendations for standardized procedures for the online and offline measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide. *Am. J. Respir. Crit. Care Med.* **2005**, *171*, 912–930. [[CrossRef](#)] [[PubMed](#)]
12. Marthin, J.K.; Nielsen, K.G. Hand-held tidal breathing nasal nitric oxide measurement—A promising targeted case-finding tool for the diagnosis of primary ciliary dyskinesia. *PLoS ONE* **2013**, *8*, e57262. [[CrossRef](#)] [[PubMed](#)]
13. Mateos-Corral, D.; Coombs, R.; Grasemann, H.; Ratjen, F.; Dell, S.D. Diagnostic value of nasal nitric oxide measured with non-velum closure techniques for children with primary ciliary dyskinesia. *J. Pediatr.* **2011**, *159*, 420–424. [[CrossRef](#)] [[PubMed](#)]
14. Harris, A.; Bhullar, E.; Gove, K.; Joslin, R.; Pelling, J.; Evans, H.J.; Walker, W.T.; Lucas, J.S. Validation of a portable nitric oxide analyzer for screening in primary ciliary dyskinesias. *BMC Pulm. Med.* **2014**, *14*, 18. [[CrossRef](#)] [[PubMed](#)]
15. Beydon, N.; Chambellan, A.; Alberti, C.; de Blic, J.; Clément, A.; Escudier, E.; Le Bourgeois, M. Technical and practical issues for tidal breathing measurements of nasal nitric oxide in children. *Pediatr. Pulmonol.* **2015**, *50*, 1374–1382. [[CrossRef](#)] [[PubMed](#)]
16. Dweik, R.A.; Boggs, P.B.; Erzurum, S.C.; Irvin, C.G.; Leigh, M.W.; Lundberg, J.O.; Olin, A.C.; Plummer, A.L.; Taylor, D.R.; on behalf of the American Thoracic Society Committee on Interpretation of Exhaled Nitric Oxide Levels (F_{ENO}) for Clinical Applications. An official ATS clinical practice guideline: Interpretation of exhaled nitric oxide levels (F_{ENO}) for clinical applications. *Am. J. Respir. Crit. Care Med.* **2011**, *184*, 602–615. [[CrossRef](#)] [[PubMed](#)]
17. Baraldi, E.; Scollo, M.; Zaramella, C.; Zanconato, S.; Zacchello, F. A simple flow-driven method for online measurement of exhaled NO starting at the age of 4 to 5 years. *Am. J. Respir. Crit. Care Med.* **2000**, *162*, 1828–1832. [[CrossRef](#)]
18. Baraldi, E.; de Jongste, J.C. European Respiratory Society/American Thoracic Society (ERS/ATS) Task Force. Measurement of exhaled nitric oxide in children, 2001. *Eur. Respir. J.* **2002**, *20*, 223–237.
19. Franklin, P.J.; Turner, S.W.; Mutch, R.C.; Stick, S.M. Comparison of single-breath and tidal breathing exhaled nitric oxide levels in infants. *Eur. Respir. J.* **2004**, *23*, 369–372. [[CrossRef](#)]
20. Silkoff, P.E.; Stevens, A.; Pak, J.; Bucher-Bartelson, B.; Martin, R.J. A method for the standardized offline collection of exhaled nitric oxide. *Chest* **1999**, *116*, 754–759. [[CrossRef](#)]

21. Van der Heijden, H.H.; Brouwer, M.L.; Hoekstra, F.; van der Pol, P.; Merkus, P.J. Reference values of exhaled nitric oxide in healthy children 1–5 years using off-line tidal breathing. *Pediatr. Pulmonol.* **2014**, *49*, 291–295. [[CrossRef](#)] [[PubMed](#)]
22. Pijnenburg, M.W. The Role of FeNO in Predicting Asthma. *Front. Pediatr.* **2019**, *7*, 41. [[CrossRef](#)] [[PubMed](#)]
23. Heijkenskjöld-Rentzhog, C.; Kalm-Stephens, P.; Nordvall, L.; Malinovski, A.; Alving, K. New method for single-breath fraction of exhaled nitric oxide measurement with improved feasibility in preschool children with asthma. *Pediatr. Allergy Immunol.* **2015**, *26*, 662–667. [[CrossRef](#)]
24. Van Mastrigt, E.; de Groot, R.C.A.; van Kesteren, H.W.; Vink, A.T.J.; de Jongste, J.C.; Pijnenburg, M.W.H. Tidal breathing FeNO measurements: A new algorithm. *Pediatr. Pulmonol.* **2014**, *49*, 15–20. [[CrossRef](#)]
25. Magit, A. Pediatric rhinosinusitis. *Otolaryngol. Clin. N. Am.* **2014**, *47*, 733–746. [[CrossRef](#)]
26. Alving, K.; Weitzberg, E.; Lundberg, J.M. Increased amount of nitric oxide in exhaled air of asthmatics. *Eur. Respir. J.* **1993**, *6*, 1368–1370.
27. Lundberg, J.O.; Weitzberg, E. Nasal nitric oxide in man. *Thorax* **1999**, *54*, 947–952. [[CrossRef](#)]
28. Lundberg, J.O.; Farkas-Szallasi, T.; Weitzberg, E.; Rinder, J.; Lidholm, J.; Anggård, A.; Hökfelt, T.; Lundberg, J.M.; Alving, K. High nitric oxide production in human paranasal sinuses. *Nat. Med.* **1995**, *1*, 370–373. [[CrossRef](#)]
29. Schairer, D.O.; Chouake, J.S.; Nosanchuk, J.D.; Friedman, A.J. The potential of nitric oxide releasing therapies as antimicrobial agents. *Virulence* **2012**, *3*, 271–279. [[CrossRef](#)]
30. Lindberg, S.; Cervin, A.; Runer, T. Low levels of nasal nitric oxide (NO) correlate to impaired mucociliary function in the upper airways. *Acta Oto-Laryngol.* **1997**, *117*, 728–734. [[CrossRef](#)]
31. Holden, W.E.; Wilkins, J.P.; Harris, M.; Milczuk, H.A.; Giraud, G.D. Temperature conditioning of nasal air: Effects of vasoactive agents and involvement of nitric oxide. *J. Appl. Physiol.* **1999**, *87*, 1260–1265. [[CrossRef](#)] [[PubMed](#)]
32. Gerlach, H.; Rossaint, R.; Pappert, D.; Knorr, M.; Falke, K.J. Autoinhalation of nitric oxide after endogenous synthesis in nasopharynx. *Lancet* **1994**, *343*, 518–519. [[CrossRef](#)]
33. Lundberg, J. Airborne nitric oxide: Inflammatory marker and aerocrine messenger in man. *Acta Physiol. Scand.* **1996**, *157*, 4–27. [[CrossRef](#)]
34. Baraldi, E.; Azzolin, N.M.; Biban, P.; Zacchello, F. Effect of antibiotic therapy on nasal nitric oxide concentration in children with acute sinusitis. *Am. J. Respir. Crit. Care Med.* **1997**, *155*, 1680–1683. [[CrossRef](#)]
35. Colantonio, D.; Brouillette, L.; Parikh, A.; Scadding, G.K. Paradoxical low nasal nitric oxide in nasal polyposis. *Clin. Exp. Allergy* **2002**, *32*, 698–701. [[CrossRef](#)]
36. Ragab, S.M.; Lund, V.J.; Saleh, H.A.; Scadding, G. Nasal nitric oxide in objective evaluation of chronic rhinosinusitis therapy. *Allergy* **2006**, *61*, 717–724. [[CrossRef](#)]
37. Arnal, J.F.; Flores, P.; Rami, J.; Murriss-Espin, M.; Bremont, F.; Pasto, I.; Aguilla, M.; Serrano, E.; Didier, A. Nasal nitric oxide concentration in paranasal sinus inflammatory diseases. *Eur. Respir. J.* **1999**, *13*, 307–312. [[CrossRef](#)]
38. Rouby, J.-J. The nose, nitric oxide, and paranasal sinuses: The outpost of pulmonary anti-infectious defenses? *Am. J. Respir. Crit. Care Med.* **2003**, *168*, 265–266. [[CrossRef](#)]
39. Phillips, P.S.; Sacks, R.; Marcells, G.N.; Cohen, N.A.; Harvey, R.J. Nasal nitric oxide and sinonasal disease: A systematic review of published evidence. *Otolaryngol. Head Neck Surg.* **2011**, *144*, 159–169. [[CrossRef](#)]
40. Kobayashi, Y.; Asako, M.; Ooka, H.; Kanda, A.; Tomoda, K.; Yasuba, H. Residual exhaled nitric oxide elevation in asthmatics is associated with eosinophilic chronic rhinosinusitis. *J. Asthma* **2015**, *52*, 1060–1064. [[CrossRef](#)] [[PubMed](#)]
41. Zhang, J.; Sun, Y.; Liu, M.; Sun, C.; Tian, L. Predictive and Diagnostic Value of Fractional Exhaled Nitric Oxide in Patients with Chronic Rhinosinusitis. *Med. Sci. Monit. Int. Med. J. Exp. Clin. Res.* **2019**, *25*, 150–156. [[CrossRef](#)] [[PubMed](#)]
42. Jeong, J.H.; Yoo, H.S.; Lee, S.H.; Kim, K.R.; Yoon, H.J.; Kim, S.H. Nasal and exhaled nitric oxide in chronic rhinosinusitis with polyps. *Am. J. Rhinol. Allergy* **2014**, *28*, e11–e16. [[CrossRef](#)] [[PubMed](#)]
43. Takeno, S.; Taruya, T.; Ueda, T.; Noda, N.; Hirakawa, K. Increased exhaled nitric oxide and its oxidation metabolism in eosinophilic chronic rhinosinusitis. *Auris Nasus Larynx* **2013**, *40*, 458–464. [[CrossRef](#)]
44. Kambara, R.; Minami, T.; Akazawa, H.; Tsuji, F.; Sasaki, T.; Inohara, H.; Horii, A. Lower Airway Inflammation in Eosinophilic Chronic Rhinosinusitis as Determined by Exhaled Nitric Oxide. *Int. Arch. Allergy Immunol.* **2017**, *173*, 225–232. [[CrossRef](#)]

45. Nakagami, Y.; Favoreto, S.; Zhen, G.; Park, S.-W.; Nguyenvu, L.T.; Kuperman, D.A.; Dolganov, G.M.; Huang, X.; Boushey, H.A.; Avila, P.C.; et al. The epithelial anion transporter pendrin is induced by allergy and rhinovirus infection, regulates airway surface liquid, and increases airway reactivity and inflammation in an asthma model. *J. Immunol.* **2008**, *181*, 2203–2210. [[CrossRef](#)]
46. Nakao, I.; Kanaji, S.; Ohta, S.; Matsushita, H.; Arima, K.; Yuyama, N.; Yamaya, M.; Nakayama, K.; Kubo, H.; Watanabe, M.; et al. Identification of pendrin as a common mediator for mucus production in bronchial asthma and chronic obstructive pulmonary disease. *J. Immunol.* **2008**, *180*, 6262–6269. [[CrossRef](#)]
47. Garnett, J.P.; Hickman, E.; Burrows, R.; Hegyi, P.; Tiszlavicz, L.; Cuthbert, A.W.; Fong, P.; Gray, M.A. Novel role for pendrin in orchestrating bicarbonate secretion in cystic fibrosis transmembrane conductance regulator (CFTR)-expressing airway serous cells. *J. Biol. Chem.* **2011**, *286*, 41069–41082. [[CrossRef](#)]
48. Ishida, A.; Ohta, N.; Suzuki, Y.; Kakehata, S.; Okubo, K.; Ikeda, H.; Shiraiishi, H.; Izuhara, K. Expression of pendrin and periostin in allergic rhinitis and chronic rhinosinusitis. *Allergol. Int.* **2012**, *61*, 589–595. [[CrossRef](#)]
49. Seshadri, S.; Lu, X.; Purkey, M.R.; Homma, T.; Choi, A.W.; Carter, R.; Suh, L.; Norton, J.; Harris, K.E.; Conley, D.B.; et al. Increased expression of the epithelial anion transporter pendrin/SLC26A4 in nasal polyps of patients with chronic rhinosinusitis. *J. Allergy Clin. Immunol.* **2015**, *136*, 1548–1558.e7. [[CrossRef](#)]
50. Jia, G.; Erickson, R.W.; Choy, D.F.; Mosesova, S.; Wu, L.C.; Solberg, O.D.; Shikotra, A.; Carter, R.; Audusseau, S.; Hamid, Q.; et al. Periostin is a systemic biomarker of eosinophilic airway inflammation in asthmatic patients. *J. Allergy Clin. Immunol.* **2012**, *130*, 647–654.e10. [[CrossRef](#)]
51. Loutsios, C.; Farahi, N.; Porter, L.; Lok, L.S.C.; Peters, A.M.; Condliffe, A.M.; Chilvers, E.R. Biomarkers of eosinophilic inflammation in asthma. *Expert Rev. Respir. Med.* **2014**, *8*, 143–150. [[CrossRef](#)] [[PubMed](#)]
52. Takayama, G.; Arima, K.; Kanaji, T.; Toda, S.; Tanaka, H.; Shoji, S.; McKenzie, A.N.; Nagai, H.; Hotokebuchi, T.; Izuhara, K. Periostin: A novel component of subepithelial fibrosis of bronchial asthma downstream of IL-4 and IL-13 signals. *J. Allergy Clin. Immunol.* **2006**, *118*, 98–104. [[CrossRef](#)] [[PubMed](#)]
53. Ebenezer, J.A.; Christensen, J.M.; Oliver, B.G.; Oliver, R.A.; Tjin, G.; Ho, J.; Habib, A.R.; Rimmer, J.; Sacks, R.; Harvey, R.J. Periostin as a marker of mucosal remodelling in chronic rhinosinusitis. *Rhinology* **2017**, *55*, 234–241. [[CrossRef](#)] [[PubMed](#)]
54. Stankovic, K.M.; Goldsztein, H.; Reh, D.D.; Platt, M.P.; Metson, R. Gene expression profiling of nasal polyps associated with chronic sinusitis and aspirin-sensitive asthma. *Laryngoscope* **2008**, *118*, 881–889. [[CrossRef](#)]
55. Maxfield, A.Z.; Landegger, L.D.; Brook, C.D.; Lehmann, A.E.; Campbell, A.P.; Bergmark, R.W.; Stankovic, K.M.; Metson, R. Periostin as a Biomarker for Nasal Polyps in Chronic Rhinosinusitis. *Otolaryngol. Head Neck Surg.* **2018**, *158*, 181–186. [[CrossRef](#)]
56. Xu, M.; Chen, D.; Zhou, H.; Zhang, W.; Xu, J.; Chen, L. The Role of Periostin in the Occurrence and Progression of Eosinophilic Chronic Sinusitis with Nasal Polyps. *Sci. Rep.* **2017**, *7*, 9479. [[CrossRef](#)]
57. Xu, M.; Zhang, W.; Chen, D.; Zhou, H.; Chen, L. Diagnostic significance of serum periostin in eosinophilic chronic sinusitis with nasal polyps. *Acta Otolaryngol.* **2018**, *138*, 387–391. [[CrossRef](#)]
58. Jonstam, K.; Westman, M.; Holtappels, G.; Holweg, C.T.J.; Bachert, C. Serum periostin, IgE, and SE-IgE can be used as biomarkers to identify moderate to severe chronic rhinosinusitis with nasal polyps. *J. Allergy Clin. Immunol.* **2017**, *140*, 1705–1708.e3. [[CrossRef](#)]
59. Papi, A.; Brightling, C.; Pedersen, S.E.; Reddel, H.K. Asthma. *Lancet* **2018**, *391*, 783–800. [[CrossRef](#)]
60. Landgraf-Rauf, K.; Anselm, B.; Schaub, B. The puzzle of immune phenotypes of childhood asthma. *Mol. Cell Pediatr.* **2016**, *3*, 27. [[CrossRef](#)]
61. Pavord, I.D.; Beasley, R.; Agusti, A.; Anderson, G.P.; Bel, E.; Brusselle, G.; Cullinan, P.; Custovic, A.; Ducharme, F.M.; Fahy, J.V.; et al. After asthma: Redefining airways diseases. *Lancet* **2018**, *391*, 350–400. [[CrossRef](#)]
62. Borland, C.; Cox, Y.; Higenbottam, T. Measurement of exhaled nitric oxide in man. *Thorax* **1993**, *48*, 1160–1162. [[CrossRef](#)] [[PubMed](#)]
63. Nelson, B.V.; Sears, S.; Woods, J.; Ling, C.Y.; Hunt, J.; Clapper, L.M.; Gaston, B. Expired nitric oxide as a marker for childhood asthma. *J. Pediatr.* **1997**, *130*, 423–427. [[CrossRef](#)]
64. Baraldi, E.; Azzolin, N.M.; Zanconato, S.; Dario, C.; Zacchello, F. Corticosteroids decrease exhaled nitric oxide in children with acute asthma. *J. Pediatr.* **1997**, *131*, 381–385. [[CrossRef](#)]
65. Mahr, T.A.; Malka, J.; Spahn, J.D. Inflammometry in pediatric asthma: A review of fractional exhaled nitric oxide in clinical practice. *Allergy Asthma Proc.* **2013**, *34*, 210–219. [[CrossRef](#)]

66. Pijnenburg, M.W.H.; De Jongste, J.C. Exhaled nitric oxide in childhood asthma: A review. *Clin. Exp. Allergy* **2008**, *38*, 246–259. [[CrossRef](#)]
67. Fleming, L.; Tsartsali, L.; Wilson, N.; Regamey, N.; Bush, A. Longitudinal relationship between sputum eosinophils and exhaled nitric oxide in children with asthma. *Am. J. Respir. Crit. Care Med.* **2013**, *188*, 400–402. [[CrossRef](#)]
68. Karrasch, S.; Linde, K.; Rücker, G.; Sommer, H.; Karsch-Völk, M.; Kleijnen, J.; Jörres, R.A.; Schneider, A. Accuracy of $FeNO$ for diagnosing asthma: A systematic review. *Thorax* **2017**, *72*, 109–116. [[CrossRef](#)]
69. Taylor, D.R.; Pijnenburg, M.W.; Smith, A.D.; De Jongste, J.C. Exhaled nitric oxide measurements: Clinical application and interpretation. *Thorax* **2006**, *61*, 817–827. [[CrossRef](#)]
70. Moeller, A.; Carlsen, K.-H.; Sly, P.D.; Baraldi, E.; Piacentini, G.; Pavord, I.; Lex, C.; Saglani, S.; on behalf of the ERS Task Force Monitoring Asthma in Children. Monitoring asthma in childhood: Lung function, bronchial responsiveness and inflammation. *Eur. Respir. Rev.* **2015**, *24*, 204–215. [[CrossRef](#)]
71. Baraldi, E.; Dario, C.; Ongaro, R.; Scollo, M.; Azzolin, N.M.; Panza, N.; Paganini, N.; Zacchello, F. Exhaled nitric oxide concentrations during treatment of wheezing exacerbation in infants and young children. *Am. J. Respir. Crit. Care Med.* **1999**, *159*, 1284–1288. [[CrossRef](#)] [[PubMed](#)]
72. Sayão, L.B.; de Britto, M.C.A.; Burity, E.; Rattes, C.; Reinaux, C.M.A.; Fink, J.; Dornelas de Andrade, A. Exhaled nitric oxide as a diagnostic tool for wheezing in preschool children: A diagnostic accuracy study. *Respir. Med.* **2016**, *113*, 15–21. [[CrossRef](#)] [[PubMed](#)]
73. Soh, J.E.; Kim, K.-M.; Kwon, J.-W.; Kim, H.Y.; Seo, J.-H.; Kim, H.-B.; Lee, S.Y.; Jang, G.C.; Song, D.J.; Kim, W.K.; et al. Recurrent wheeze and its relationship with lung function and airway inflammation in preschool children: A cross-sectional study in South Korea. *BMJ Open* **2017**, *7*, e018010. [[CrossRef](#)] [[PubMed](#)]
74. Moeller, A.; Diefenbacher, C.; Lehmann, A.; Rochat, M.; Brooks-Wildhaber, J.; Hall, G.L.; Wildhaber, J.H. Exhaled nitric oxide distinguishes between subgroups of preschool children with respiratory symptoms. *J. Allergy Clin. Immunol.* **2008**, *121*, 705–709. [[CrossRef](#)]
75. Castro-Rodriguez, J.A.; Sardón, O.; Pérez-Yarza, E.G.; Korta, J.; Aldasoro, A.; Corcuera, P.; Mintegui, J. Young infants with recurrent wheezing and positive asthma predictive index have higher levels of exhaled nitric oxide. *J. Asthma* **2013**, *50*, 162–165. [[CrossRef](#)]
76. Oh, M.-A.; Shim, J.Y.; Jung, Y.-H.; Seo, J.-H.; Young Kim, H.; Kwon, J.-W.; Kim, B.J.; Kim, H.B.; Kim, W.K.; Lee, S.Y.; et al. Fraction of exhaled nitric oxide and wheezing phenotypes in preschool children. *Pediatr. Pulmonol.* **2013**, *48*, 563–570. [[CrossRef](#)]
77. Singer, F.; Luchsinger, I.; Inci, D.; Knauer, N.; Latzin, P.; Wildhaber, J.H.; Moeller, A. Exhaled nitric oxide in symptomatic children at preschool age predicts later asthma. *Allergy* **2013**, *68*, 531–538. [[CrossRef](#)]
78. Caudri, D.; Wijga, A.H.; Hoekstra, M.O.; Kerkhof, M.; Koppelman, G.H.; Brunekreef, B.; Smit, H.A.; de Jongste, J.C. Prediction of asthma in symptomatic preschool children using exhaled nitric oxide, Rint and specific IgE. *Thorax* **2010**, *65*, 801–807. [[CrossRef](#)]
79. Elliott, M.; Heltshe, S.L.; Stamey, D.C.; Cochrane, E.S.; Redding, G.J.; Debley, J.S. Exhaled nitric oxide predicts persistence of wheezing, exacerbations, and decline in lung function in wheezy infants and toddlers. *Clin. Exp. Allergy* **2013**, *43*, 1351–1361. [[CrossRef](#)]
80. Debley, J.S.; Stamey, D.C.; Cochrane, E.S.; Gama, K.L.; Redding, G.J. Exhaled nitric oxide, lung function, and exacerbations in wheezy infants and toddlers. *J. Allergy Clin. Immunol.* **2010**, *125*, 1228–1234.e13. [[CrossRef](#)]
81. Linn, W.S.; Rappaport, E.B.; Berhane, K.T.; Bastain, T.M.; Avol, E.L.; Gilliland, F.D. Exhaled nitric oxide in a population-based study of southern California schoolchildren. *Respir. Res.* **2009**, *10*, 28. [[CrossRef](#)] [[PubMed](#)]
82. Di Cara, G.; Marcucci, F.; Palomba, A.; Milioni, M.; Pecoraro, L.; Ciprandi, G.; Buttafava, S.; Frati, F.; Verrotti, A. Exhaled nitric oxide in children with allergic rhinitis: A potential biomarker of asthma development. *Pediatr. Allergy Immunol.* **2015**, *26*, 85–87. [[CrossRef](#)] [[PubMed](#)]
83. Horváth, I.; Barnes, P.J.; Loukides, S.; Sterk, P.J.; Högman, M.; Olin, A.-C.; Amann, A.; Antus, B.; Baraldi, E.; Bikov, A.; et al. A European Respiratory Society technical standard: Exhaled biomarkers in lung disease. *Eur. Respir. J.* **2017**, *49*, 1600965. [[CrossRef](#)]
84. Moeller, A.; Franklin, P.; Hall, G.L.; Turner, S.; Straub, D.; Wildhaber, J.H.; Stick, S.M. Inhaled fluticasone dipropionate decreases levels of nitric oxide in recurrently wheezy infants. *Pediatr. Pulmonol.* **2004**, *38*, 250–255. [[CrossRef](#)]

85. Petsky, H.L.; Cates, C.J.; Kew, K.M.; Chang, A.B. Tailoring asthma treatment on eosinophilic markers (exhaled nitric oxide or sputum eosinophils): A systematic review and meta-analysis. *Thorax* **2018**, *73*, 1110–1119. [[CrossRef](#)]
86. NICE. Asthma: Diagnosis, Monitoring and Chronic Asthma Management. Guidance and Guidelines. Available online: <https://www.nice.org.uk/guidance/ng80> (accessed on 3 July 2018).
87. SIGN 158 British Guideline on the Management of Asthma. Available online: <https://www.sign.ac.uk/sign-158-british-guideline-on-the-management-of-asthma.html> (accessed on 19 August 2019).
88. 2019 GINA Main Report. Global Initiative for Asthma—GINA. Available online: <https://ginasthma.org/gina-reports/> (accessed on 23 July 2019).
89. Fingleton, J.; Braithwaite, I.; Travers, J.; Bowles, D.; Strik, R.; Siebers, R.; Holweg, C.; Matthews, J.; Weatherall, M.; Beasley, R. Serum periostin in obstructive airways disease. *Eur. Respir. J.* **2016**, *47*, 1383–1391. [[CrossRef](#)]
90. Kanemitsu, Y.; Matsumoto, H.; Izuhara, K.; Tohda, Y.; Kita, H.; Horiguchi, T.; Kuwabara, K.; Tomii, K.; Otsuka, K.; Fujimura, M.; et al. Increased periostin associates with greater airflow limitation in patients receiving inhaled corticosteroids. *J. Allergy Clin. Immunol.* **2013**, *132*, 305–312.e3. [[CrossRef](#)]
91. Matsusaka, M.; Kabata, H.; Fukunaga, K.; Suzuki, Y.; Masaki, K.; Mochimaru, T.; Sakamaki, F.; Oyamada, Y.; Inoue, T.; Oguma, T.; et al. Phenotype of asthma related with high serum periostin levels. *Allergol. Int.* **2015**, *64*, 175–180. [[CrossRef](#)]
92. Sidhu, S.S.; Yuan, S.; Innes, A.L.; Kerr, S.; Woodruff, P.G.; Hou, L.; Muller, S.J.; Fahy, J.V. Roles of epithelial cell-derived periostin in TGF-beta activation, collagen production, and collagen gel elasticity in asthma. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 14170–14175. [[CrossRef](#)]
93. James, A.; Hedlin, G. Biomarkers for the Phenotyping and Monitoring of Asthma in Children. *Curr. Treat. Options Allergy* **2016**, *3*, 439–452. [[CrossRef](#)]
94. Anderson, H.M.; Lemanske, R.F.; Arron, J.R.; Holweg, C.T.J.; Rajamanickam, V.; Gangnon, R.E.; Gern, J.E.; Jackson, D.J. Relationships among aeroallergen sensitization, peripheral blood eosinophils, and periostin in pediatric asthma development. *J. Allergy Clin. Immunol.* **2017**, *139*, 790–796. [[CrossRef](#)] [[PubMed](#)]
95. Inoue, T.; Akashi, K.; Watanabe, M.; Ikeda, Y.; Ashizuka, S.; Motoki, T.; Suzuki, R.; Sagara, N.; Yanagida, N.; Sato, S.; et al. Periostin as a biomarker for the diagnosis of pediatric asthma. *Pediatr. Allergy Immunol.* **2016**, *27*, 521–526. [[CrossRef](#)] [[PubMed](#)]
96. Licari, A.; Castagnoli, R.; Brambilla, I.; Marseglia, A.; Tosca, M.A.; Marseglia, G.L.; Ciprandi, G. Asthma Endotyping and Biomarkers in Childhood Asthma. *Pediatr. Allergy Immunol. Pulmonol.* **2018**, *1*, 3144–3155. [[CrossRef](#)] [[PubMed](#)]
97. Davis, M.D.; Montpetit, A.J. Exhaled Breath Condensate: An Update. *Immunol. Allergy Clin. N. Am.* **2018**, *38*, 667–678. [[CrossRef](#)]
98. Moschino, L.; Zanconato, S.; Bozzetto, S.; Baraldi, E.; Carraro, S. Childhood asthma biomarkers: Present knowledge and future steps. *Paediatr. Respir. Rev.* **2015**, *16*, 205–212. [[CrossRef](#)]
99. Ferraro, V.; Carraro, S.; Bozzetto, S.; Zanconato, S.; Baraldi, E. Exhaled biomarkers in childhood asthma: Old and new approaches. *Asthma Res. Pract.* **2018**, *4*, 9. [[CrossRef](#)]
100. Horváth, I.; Hunt, J.; Barnes, P.J.; Alving, K.; Antczak, A.; Baraldi, E.; Becher, G.; van Beurden, W.J.; Corradi, M.; Dekhuijzen, R.; et al. Exhaled breath condensate: Methodological recommendations and unresolved questions. *Eur. Respir. J.* **2005**, *26*, 523–548. [[CrossRef](#)]
101. Thomas, P.S.; Lowe, A.J.; Samarasinghe, P.; Lodge, C.J.; Huang, Y.; Abramson, M.J.; Dharmage, S.C.; Jaffe, A. Exhaled breath condensate in pediatric asthma: Promising new advance or pouring cold water on a lot of hot air? A systematic review. *Pediatr. Pulmonol.* **2013**, *48*, 419–442. [[CrossRef](#)]
102. Morton, J.; Henry, R.L.; Thomas, P.S. Exhaled breath condensate nitrite/nitrate and pH in relation to pediatric asthma control and exhaled nitric oxide. *Pediatr. Pulmonol.* **2006**, *41*, 929–936.
103. Brunetti, L.; Francavilla, R.; Tesse, R.; Strippoli, A.; Polimeno, L.; Loforese, A.; Miniello, V.L.; Armenio, L. Exhaled breath condensate pH measurement in children with asthma, allergic rhinitis and atopic dermatitis. *Pediatr. Allergy Immunol.* **2006**, *17*, 422–427. [[CrossRef](#)]
104. Montuschi, P. LC/MS/MS analysis of leukotriene B4 and other eicosanoids in exhaled breath condensate for assessing lung inflammation. *J. Chromatogr. B* **2009**, *877*, 1272–1280. [[CrossRef](#)] [[PubMed](#)]
105. Montuschi, P.; Barnes, P.J. Exhaled leukotrienes and prostaglandins in asthma. *J. Allergy Clin. Immunol.* **2002**, *109*, 615–620. [[CrossRef](#)] [[PubMed](#)]

106. Wan, G.-H.; Yan, D.-C.; Tseng, H.-Y.; Tung, T.-H.; Lin, S.-J.; Lin, Y.-W. Cysteinyl leukotriene levels correlate with 8-isoprostane levels in exhaled breath condensates of atopic and healthy children. *Pediatr. Res.* **2013**, *74*, 584–591. [[CrossRef](#)]
107. Baraldi, E.; Carraro, S.; Alinovi, R.; Pesci, A.; Ghio, L.; Bodini, A.; Piacentini, G.; Zacchello, F.; Zanconato, S. Cysteinyl leukotrienes and 8-isoprostane in exhaled breath condensate of children with asthma exacerbations. *Thorax* **2003**, *58*, 505–509. [[CrossRef](#)]
108. Samitas, K.; Chorianopoulos, D.; Vittorakis, S.; Zervas, E.; Economidou, E.; Papatheodorou, G.; Loukides, S.; Gaga, M. Exhaled cysteinyl-leukotrienes and 8-isoprostane in patients with asthma and their relation to clinical severity. *Respir. Med.* **2009**, *103*, 750–756. [[CrossRef](#)]
109. Bodini, A.; Peroni, D.; Vicentini, L.; Loiacono, A.; Baraldi, E.; Ghio, L.; Corradi, M.; Alinovi, R.; Boner, A.L.; Piacentini, G.L. Exhaled breath condensate eicosanoids and sputum eosinophils in asthmatic children: A pilot study. *Pediatr. Allergy Immunol.* **2004**, *15*, 26–31. [[CrossRef](#)]
110. Teng, Y.; Sun, P.; Zhang, J.; Yu, R.; Bai, J.; Yao, X.; Huang, M.; Adcock, I.M.; Barnes, P.J. Hydrogen peroxide in exhaled breath condensate in patients with asthma: A promising biomarker? *Chest* **2011**, *140*, 108–116. [[CrossRef](#)]
111. Jöbssis, Q.; Raatgeep, H.C.; Hermans, P.W.; de Jongste, J.C. Hydrogen peroxide in exhaled air is increased in stable asthmatic children. *Eur. Respir. J.* **1997**, *10*, 519–521.
112. Baraldi, E.; Giordano, G.; Pasquale, M.F.; Carraro, S.; Mardegan, A.; Bonetto, G.; Bastardo, C.; Zacchello, F.; Zanconato, S. 3-Nitrotyrosine, a marker of nitrosative stress, is increased in breath condensate of allergic asthmatic children. *Allergy* **2006**, *61*, 90–96. [[CrossRef](#)]
113. Formanek, W.; Inci, D.; Lauener, R.P.; Wildhaber, J.H.; Frey, U.; Hall, G.L. Elevated nitrite in breath condensates of children with respiratory disease. *Eur. Respir. J.* **2002**, *19*, 487–491. [[CrossRef](#)]
114. Wheelock, C.E.; Goss, V.M.; Balgoma, D.; Nicholas, B.; Brandsma, J.; Skipp, P.J.; Snowden, S.; Burg, D.; D'Amico, A.; Horvath, I.; et al. Application of 'omics technologies to biomarker discovery in inflammatory lung diseases. *Eur. Respir. J.* **2013**, *42*, 802–825. [[CrossRef](#)] [[PubMed](#)]
115. Bush, A. Translating Asthma: Dissecting the Role of Metabolomics, Genomics and Personalized Medicine. *Indian J. Pediatr.* **2018**, *85*, 643–650. [[CrossRef](#)] [[PubMed](#)]
116. Lin, J.-L.; Bonnicksen, M.H.; Nogeh, E.U.; Raftery, M.J.; Thomas, P.S. Proteomics in detection and monitoring of asthma and smoking-related lung diseases. *Expert Rev. Proteom.* **2010**, *7*, 361–372. [[CrossRef](#)] [[PubMed](#)]
117. Wiktorowicz, J.E.; Jamaluddin, M. Proteomic analysis of the asthmatic airway. *Adv. Exp. Med. Biol.* **2014**, *795*, 221–232.
118. Lacombe, M.; Marie-Desvergne, C.; Combes, F.; Kraut, A.; Bruley, C.; Vandebrouck, Y.; Chamel Mossuz, V.; Couté, Y.; Brun, V. Proteomic characterization of human exhaled breath condensate. *J. Breath Res.* **2018**, *12*, 021001. [[CrossRef](#)]
119. Muccilli, V.; Saletti, R.; Cunsolo, V.; Ho, J.; Gili, E.; Conte, E.; Sichili, S.; Vancheri, C.; Foti, S. Protein profile of exhaled breath condensate determined by high resolution mass spectrometry. *J. Pharm. Biomed. Anal.* **2015**, *105*, 134–149. [[CrossRef](#)]
120. Bloemen, K.; Van Den Heuvel, R.; Govarts, E.; Hooyberghs, J.; Nelen, V.; Witters, E.; Desager, K.; Schoeters, G. A new approach to study exhaled proteins as potential biomarkers for asthma. *Clin. Exp. Allergy* **2011**, *41*, 346–356. [[CrossRef](#)]
121. Turi, K.N.; Romick-Rosendale, L.; Ryckman, K.K.; Hartert, T.V. A review of metabolomics approaches and their application in identifying causal pathways of childhood asthma. *J. Allergy Clin. Immunol.* **2018**, *141*, 1191–1201. [[CrossRef](#)]
122. Carraro, S.; Giordano, G.; Reniero, F.; Perilongo, G.; Baraldi, E. Metabolomics: A new frontier for research in pediatrics. *J. Pediatr.* **2009**, *154*, 638–644. [[CrossRef](#)]
123. Carraro, S.; Rezzi, S.; Reniero, F.; Héberger, K.; Giordano, G.; Zanconato, S.; Guillou, C.; Baraldi, E. Metabolomics applied to exhaled breath condensate in childhood asthma. *Am. J. Respir. Crit. Care Med.* **2007**, *175*, 986–990. [[CrossRef](#)]
124. Carraro, S.; Giordano, G.; Reniero, F.; Carpi, D.; Stocchero, M.; Sterk, P.J.; Baraldi, E. Asthma severity in childhood and metabolomic profiling of breath condensate. *Allergy* **2013**, *68*, 110–117. [[CrossRef](#)] [[PubMed](#)]
125. Konstantinidi, E.M.; Lappas, A.S.; Tzortzi, A.S.; Behrakis, P.K. Exhaled Breath Condensate: Technical and Diagnostic Aspects. *Sci. World J.* **2015**, *2015*, 435160. [[CrossRef](#)] [[PubMed](#)]

126. Bannier, M.A.; Rosias, P.P.; Jöbsis, Q.; Dompeling, E. Exhaled Breath Condensate in Childhood Asthma: A Review and Current Perspective. *Front. Pediatr.* **2019**, *7*, 150. [[CrossRef](#)] [[PubMed](#)]
127. Neerincx, A.H.; Vijverberg, S.J.H.; Bos, L.D.J.; Brinkman, P.; van der Schee, M.P.; de Vries, R.; Sterk, P.J.; Maitland-van der Zee, A.H. Breathomics from exhaled volatile organic compounds in pediatric asthma. *Pediatr. Pulmonol.* **2017**, *52*, 1616–1627. [[CrossRef](#)] [[PubMed](#)]
128. Van Mastrigt, E.; de Jongste, J.C.; Pijnenburg, M.W. The analysis of volatile organic compounds in exhaled breath and biomarkers in exhaled breath condensate in children—Clinical tools or scientific toys? *Clin. Exp. Allergy* **2015**, *45*, 1170–1188. [[CrossRef](#)] [[PubMed](#)]
129. Van der Schee, M.P.; Paff, T.; Brinkman, P.; van Aalderen, W.M.C.; Haarman, E.G.; Sterk, P.J. Breathomics in lung disease. *Chest* **2015**, *147*, 224–231. [[CrossRef](#)] [[PubMed](#)]
130. Fens, N.; van der Schee, M.P.; Brinkman, P.; Sterk, P.J. Exhaled breath analysis by electronic nose in airways disease. Established issues and key questions. *Clin. Exp. Allergy* **2013**, *43*, 705–715. [[CrossRef](#)]
131. Van Mastrigt, E.; Reyes-Reyes, A.; Brand, K.; Bhattacharya, N.; Urbach, H.P.; Stubbs, A.P.; de Jongste, J.C.; Pijnenburg, M.W. Exhaled breath profiling using broadband quantum cascade laser-based spectroscopy in healthy children and children with asthma and cystic fibrosis. *J. Breath Res.* **2016**, *10*, 026003. [[CrossRef](#)]
132. Gahleitner, F.; Guallar-Hoyas, C.; Beardsmore, C.S.; Pandya, H.C.; Thomas, C.P. Metabolomics pilot study to identify volatile organic compound markers of childhood asthma in exhaled breath. *Bioanalysis* **2013**, *5*, 2239–2247. [[CrossRef](#)]
133. Caldeira, M.; Barros, A.S.; Bilelo, M.J.; Parada, A.; Câmara, J.S.; Rocha, S.M. Profiling allergic asthma volatile metabolic patterns using a headspace-solid phase microextraction/gas chromatography based methodology. *J. Chromatogr. A* **2011**, *1218*, 3771–3780. [[CrossRef](#)]
134. Dallinga, J.W.; Robroeks, C.M.H.H.T.; van Berkel, J.J.B.N.; Moonen, E.J.C.; Godschalk, R.W.L.; Jöbsis, Q.; Dompeling, E.; Wouters, E.F.; van Schooten, F.J. Volatile organic compounds in exhaled breath as a diagnostic tool for asthma in children. *Clin. Exp. Allergy* **2010**, *40*, 68–76.
135. Caldeira, M.; Perestrelo, R.; Barros, A.S.; Bilelo, M.J.; Morête, A.; Câmara, J.S.; Rocha, S.M. Allergic asthma exhaled breath metabolome: A challenge for comprehensive two-dimensional gas chromatography. *J. Chromatogr. A* **2012**, *1254*, 87–97. [[CrossRef](#)] [[PubMed](#)]
136. Klaassen, E.M.M.; van de Kant, K.D.G.; Jöbsis, Q.; van Schayck, O.C.P.; Smolinska, A.; Dallinga, J.W.; van Schooten, F.J.; den Hartog, G.J.; de Jongste, J.C.; Rijkers, G.T.; et al. Exhaled biomarkers and gene expression at preschool age improve asthma prediction at 6 years of age. *Am. J. Respir. Crit. Care Med.* **2015**, *191*, 201–207. [[CrossRef](#)] [[PubMed](#)]
137. Smolinska, A.; Klaassen, E.M.M.; Dallinga, J.W.; van de Kant, K.D.G.; Jobsis, Q.; Moonen, E.J.C.; van Schayck, O.C.; Dompeling, E.; van Schooten, F.J. Profiling of volatile organic compounds in exhaled breath as a strategy to find early predictive signatures of asthma in children. *PLoS ONE* **2014**, *9*, e95668. [[CrossRef](#)]
138. Van Vliet, D.; Smolinska, A.; Jöbsis, Q.; Rosias, P.; Muris, J.; Dallinga, J.; Dompeling, E.; van Schooten, F.J. Can exhaled volatile organic compounds predict asthma exacerbations in children? *J. Breath Res.* **2017**, *11*, 016016. [[CrossRef](#)]
139. Robroeks, C.M.; van Berkel, J.J.; Jöbsis, Q.; van Schooten, F.-J.; Dallinga, J.W.; Wouters, E.F.; Dompeling, E. Exhaled volatile organic compounds predict exacerbations of childhood asthma in a 1-year prospective study. *Eur. Respir. J.* **2013**, *42*, 98–106. [[CrossRef](#)]
140. Van Vliet, D.; Smolinska, A.; Jöbsis, Q.; Rosias, P.P.R.; Muris, J.W.M.; Dallinga, J.W.; van Schooten, F.J.; Dompeling, E. Association between exhaled inflammatory markers and asthma control in children. *J. Breath Res.* **2016**, *10*, 016014. [[CrossRef](#)]
141. Bannier, M.A.; van de Kant, K.D.; Jöbsis, Q.; Dompeling, E. Feasibility and diagnostic accuracy of an electronic nose in children with asthma and cystic fibrosis. *J. Breath Res.* **2019**, *13*, 036009. [[CrossRef](#)]

